



## Effect of plant defense activators on white rust of mustard cv. RH- 749 under artificial epiphytotic conditions

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### Abstract

Experiment was conducted to study the effect of plant defense activators on white rust of Indian mustard cultivar RH-749 under artificial epiphytotic conditions. The size of pustule of white rust was recorded minimum in Metalaxyl 0.3% (0.32 mm) followed by Metalaxyl 0.2% (0.33 mm) during 2015-16, although, the maximum size of pustules were observed in check was 6.55 mm followed by zinc sulphate at 0.50% (5.88 mm) during 2016-17. Among the abiotic chemicals, salicylic acid was recorded significantly better among all the treatments. The number of pustules were recorded maximum in check was 7.47 and 7.53 followed by zinc sulphate at 0.50 % was 7.1 and 7.3 during 2015-16 and 2016-17, respectively. Salicylic acid 0.25% reduces 31.5% white rust disease over the control followed by calcium sulphate at 1.0% which reduces disease 24.0%. Zinc sulphate at 0.50% was found least effective abiotic chemical which reduce 6.1% disease.

**Keywords :** Epiphytotic, Indian mustard, plant defense activators, white rust

### Introduction

Plants undergo several stresses throughout their life span and respond them through numerous chemical and physical stimuli (Omid, 2010). Plants are prepared through pre-formed or constitutive mechanical and chemical barriers as well as with inducible defense systems to protect themselves from the attack of different pathogens such as bacteria, virus, fungi, nematodes and parasitic plants in their environment. Induction of defensive system in plants can be achieved or enhanced by pretreatment of plants with avirulent pathogen (biotic inducers) or chemical compounds (abiotic inducers). These biotic and abiotic inducers are known as elicitors. Elicitors are low molecular weight compounds which induces a systemic defense response in plants. These elicitors are synthesized by itself or released from polymeric precursors through infection by pathogen (Ozeretskoykaya and Vasyukova, 2002; Zhao *et al.*, 2005, Holopainen *et al.*, 2009).

Induced resistance has been exploited extensively for the management of many plant diseases (Gorlach *et al.*, 1996). Several compounds such as salicylic acid (Yalpani and Raskin, 1993), benzothiadiazoles (Narusaka *et al.*, 1999), fatty acids and oligosaccharides (Kobayashi, *et al.*, 1993), harpin protein (Wei *et al.*, 1992, De *et al.*, 1990) are effective inducers of plant defense in the host plant system. In addition, numerous microorganisms applied to the leaves or roots of plants may induce systemic or

local resistance (Liu *et al.*, 1995). Such resistance is reported to be active against several types of organisms such as fungi, bacteria and even parasitic plants (Matheron and Porchas, 2002).

Recent years, a new group of compounds that activate host defense mechanism and protect the plant against pathogens has been developed to manage crop diseases. These chemicals are known as “plant defense activators” or “plant activators” (Romero *et al.*, 2001). Salicylic acid mimics compounds such as acibenzolar s methyl, bion, phosphorous salts and micronutrient potassium salts have been used as commercial plant activators (Becot *et al.*, 2000; Macmillan *et al.*, 2000; Pajot *et al.*, 2001; Graham and Leite, 2004).

Biological plant defense inducers such as *Trichoderma viride*, *Pseudomonas*, *Bacillus*, *Serratia*, nonpathogenic strains of *Fusarium* and yeast have been developed as commercial product to combat various plant diseases (Droly *et al.*, 2002; Benhamon and Garand, 2001; Verhagen *et al.*, 2004; Howell *et al.*, 2000). *Trichoderma viride* is known as one of the most important bio-control agent that has been used extensively in agriculture. Biological plant defense inducers provide systemic resistance to plants infested by various fungal and bacterial phytopathogens. Biocontrol activity of *Trichoderma* based biocontrol agents inheres in their ability to orchestrate several biochemical pathways in diseased plants (Surekha *et al.*, 2014). Although studies explaining

biocontrol activity of *Trichoderma* against fungal pathogens are recognized, there is need for imparting the biochemical basis of disease resistance being induced by *Trichoderma*. Therefore, research investigations pertaining to induction of such systemic resistance and associated biochemical responses is necessary to understand the mechanism of biological control activity of *T. viride* and *Pseudomonas fluorescens*.

Host plant defense can be induced by the application of non-pathogenic microorganisms (Vishwanath *et al.*, 1999; Singh *et al.*, 1999) and certain abiotic activators such as salicylic acid (Spletzer and Enyedi, 1999) and amino butyric acid (Cohen, 1994; Kaur and Kolte, 2001). Host resistance is considered to be the most effective and economical method in plant disease management. Evaluation of new germplasm helps to identify some good sources of resistance that may be exploited for commercial production or for breeding of new disease resistant lines. The resistant sources help to slow down the rate of epidemic buildup. Therefore, the development of disease resistant varieties is the most effective and economical approach of integrated disease management programme.

In plants systemic resistant can be induced through biotic or abiotic plant defense inducers. Resistance to diseases can be induced systematically in plants by biological

and chemical means (Ryals *et al.*, 1994; Spletzer and Enyedi, 1999). Most commonly used chemicals inducers are salicylic acid, which appears to mimic the systemic effects of localized infection in plant system (Safari *et al.*, 2013). One of the potential management methods is the use of systemic acquired resistance to trigger host defense mechanisms, which would not involve the application of hazardous compounds to plants (Durrant and Dong, 2004). External application of salicylic acid can induce systemic acquired resistance (Hammerschmidt, 1999).

The potential of chemical inducers of systemic acquired resistance to reduce white rust disease on mustard was evaluated to explore the possibility of utilizing induced host resistance as a realistic alternative to classical fungicides in disease management.

## Materials and Methods

First spray plant defense activators in the mustard variety RH-749 when plants shows first symptom of the disease in each treatment with their respective concentration using a randomized block design. One standard chemical check Metalaxyl at 0.10, 0.20 and 0.30 per cent and one sterile distilled water check were also maintained in three replications.

Table 1: Biotic and abiotic activators and their concentration under field study

Treatments	Biotic and abiotic agents	Concentration (%)
T <sub>1</sub>	<i>Trichoderma viride</i>	1.00
T <sub>2</sub>	<i>Pseudomonas fluorescens</i>	1.00
T <sub>3</sub>	Salicylic acid	0.25
T <sub>4</sub>	Borax (Na <sub>2</sub> B <sub>4</sub> O <sub>7</sub> .10H <sub>2</sub> O)	0.50
T <sub>5</sub>	Potassium sulphate (K <sub>2</sub> SO <sub>4</sub> )	1.00
T <sub>6</sub>	Calcium sulphate (CaSO <sub>4</sub> )	1.00
T <sub>7</sub>	Metalaxyl	0.10
T <sub>8</sub>	Metalaxyl	0.20
T <sub>9</sub>	Metalaxyl	0.30
T <sub>10</sub>	Potassium chloride (KCl)	1.00
T <sub>11</sub>	Zinc sulphate (ZnSO <sub>4</sub> .7H <sub>2</sub> O)	0.50
T <sub>12</sub>	Check	-

## Observations recorded

### Size of pustule on leaves

Diameter of randomly selected five leaves was measured in mm with the help of plastic scale and average size of pustule was calculated and recorded at ten days interval.

### Number of pustules

Numbers of pustules were recorded by counting the pustules per 25 mm<sup>2</sup> leaf area of randomly selected five

leaves of plant. The observations were recorded on five leaves and average number of pustules was then calculated per 25 mm<sup>2</sup> leaf area.

### Per cent disease index on leaf

The per cent disease index on leaf due to white rust was recorded at 10 days interval up to 90 days after sowing (DAS) by using of 0-5 rating scale given by Biswas *et al.*, 2011; Tirmali and Kolte (2012).

Numerical rating	Leaf area covered by the pustules (%)
0	No symptoms
1	1-10
2	11-25
3	26-50
4	51-75
5	>75

Ratings were given as per above mentioned rating scale and white rust per cent disease index was calculated by using formula given by Wheeler (1969) and Mathur *et al.* (2013). Observations were recorded by randomly selecting twenty five leaves from each replication and were rated as per the above rating scale and per cent disease index was calculated and statistically analyzed as described by Panse and Sukhatme (1985) for analysis of variance of randomized block design in order to test the significance of experimental results.

$$\text{White Rust Index (\%)} = \frac{\text{Sum of all numerical ratings}}{\text{Number of leaves examined} \times \text{maximu grade of scale}} \times 100$$

## Results and Discussion

The size of pustule was recorded minimum in Metalaxyl 0.3% was 0.32 mm followed by Metalaxyl 0.2% (0.33 mm) in cv. RH-749 during 2015-16 (Table 2). However the different concentration of Metalaxyl was found at par with each other during 2015-16 at 70, 80 and 90 days after sowing of the crop. All the treatments were found significantly better in the comparison with the control. The maximum size of pustules was observed in check was 6.55 mm followed by zinc sulphate at 0.50% (5.88 mm) during 2016-17. Among the abiotic chemicals, salicylic acid was recorded significantly better over all the treatments. Both the biotic agents were found significantly better in comparison with the control. The

size of the pustules was enlarged from 70 to 90 days after sowing the mustard.

The number of pustules were recorded maximum in the check was 7.47 and 7.53 followed by zinc sulphate at 0.50% was 7.13 and 7.27 during 2015-16 and 2016-17, respectively (Table 3). The minimum number of pustules was found in Metalaxyl 0.2% and Metalaxyl 0.3% was 1.80. However, different concentrations of Metalaxyl were found at par in comparison with each other during 2015-16 and 2016-17 at 70, 80 and 90 days after sowing. Among the biotic agents, *T. viride* and *P. fluorescens* were found significantly better over the check except *T. viride* at 1:00% was found at par at 80 days after sowing during 2016-

Table 2: Effect of biotic and non-conventional chemicals on size of pustule on mustard cv. RH-749 under field conditions

Treatment	Concentration (%)	Size of pustules (mm)								Over all Mean	% reduction over check
		2015-16				2016-17					
		70 DAS	80 DAS	90 DAS	Mean	70 DAS	80 DAS	90 DAS	Mean		
<i>Trichoderma viride</i>	1.0	0.9	3.0	5.5	3.2	1.0	3.1	5.6	3.2	3.2	19.9
<i>Pseudomonas fluorescens</i>	1.0	0.6	2.2	4.7	2.5	0.7	2.27	4.8	2.6	2.6	36.0
Salicylic Acid	0.3	0.4	1.8	4.01	2.1	0.5	1.8	4.1	2.2	2.1	46.7
Borax	0.5	0.8	2.6	5.19	2.9	0.9	2.7	5.3	3.0	2.9	26.7
K <sub>2</sub> SO <sub>4</sub>	1.0	0.6	2.5	4.98	2.7	0.7	2.6	5.1	2.8	2.8	30.8
CaSO <sub>4</sub>	1.0	0.5	2.1	4.39	2.3	0.6	2.2	4.5	2.4	2.4	40.6
Metalaxyl 0.1%	0.1	0.4	1.6	3.40	1.8	0.4	1.7	3.5	1.9	1.8	53.9
Metalaxyl 0.2%	0.2	0.3	1.6	3.23	1.7	0.4	1.6	3.4	1.8	1.8	55.9
Metalaxyl 0.3%	0.3	0.3	1.6	3.17	1.7	0.4	1.6	3.3	1.8	1.7	56.6
KCl	1.0	0.5	2.2	4.49	2.4	0.6	2.3	4.6	2.5	2.4	38.7
ZnSO <sub>4</sub>	0.5	0.9	3.1	5.82	3.3	1.2	3.2	5.9	3.4	3.4	15.9
Check	-	1.4	4.0	6.46	3.9	1.5	4.1	6.6	4.0	4.0	0.0
SEm±	-	0.02	0.07	0.14	-	0.02	0.06	0.08	-	-	-
CD 5%	-	0.05	0.21	0.41	-	0.05	0.17	0.23	-	-	-

DAS = Days after sowing

Table 3: Effect of biotic and abiotic agents on number of white rust leaf pustule on mustard cv. RH-749 under field condition

Treatment	Concentration (%)	Number of pustules/25 mm <sup>2</sup> area												Over all	
		2015-16						2016-17						Mean	check
		70 DAS	80 DAS	90 DAS	Mean	70 DAS	80 DAS	90 DAS	Mean						
<i>Trichoderma viride</i>	1.00	3.53	5.20	7.00	5.2	3.6	5.5	7.1	5.4	5.3	9.1	5.3	9.1		
<i>Pseudomonas fluorescens</i>	1.00	2.80	4.40	5.93	4.4	2.9	4.1	6.0	4.3	4.4	25.6	4.4	25.6		
Salicylic Acid	0.25	2.40	3.40	4.93	3.6	2	3.6	5.0	3.7	3.6	37.9	3.6	37.9		
Borax	0.50	3.20	4.87	6.60	4.9	3.5	4.8	6.7	5.0	4.9	15.8	4.9	15.8		
K <sub>2</sub> SO <sub>4</sub>	1.00	3.13	4.67	6.20	4.7	3.2	4.9	6.3	4.8	4.7	19.2	4.7	19.2		
CaSO <sub>4</sub>	1.00	2.67	3.80	5.33	3.9	2.7	3.9	5.4	4.0	4.0	32.1	4.0	32.1		
Metalaxyl 0.1%	0.10	2.00	3.13	4.33	3.2	2.1	3.1	4.4	3.2	3.2	45.7	3.2	45.7		
Metalaxyl 0.2%	0.20	1.80	3.00	4.27	3.0	1.9	3.1	4.3	3.1	3.1	47.8	3.1	47.8		
Metalaxyl 0.3%	0.30	1.80	2.87	4.27	3.0	1.9	2.9	4.4	3.0	3.0	48.6	3.0	48.6		
KCl	1.00	2.93	4.00	5.47	4.1	3.0	4.1	5.5	4.2	4.2	28.8	4.2	28.8		
ZnSO <sub>4</sub>	0.50	3.87	5.27	7.1	5.4	3.7	5.3	7.3	5.4	5.4	7.4	5.4	7.4		
Check	-	4.13	5.87	7.5	5.8	4.2	5.9	7.5	5.9	5.9	0.0	5.9	0.0		
SEM±	-	0.07	0.15	0.18	-	0.07	0.15	0.18	-	-	-	-	-		
CD 5%	-	0.21	0.44	0.50	-	0.21	0.44	0.50	-	-	-	-	-		

DAS = Days after sowing

Table 4: Effect of biotic and non-conventional chemicals on disease index of white rust on mustard cv. RH-749 under field conditions

Treatment	Concentration		Disease index (%)												Over	
	(% )		2015-16						2016-17						all	over
	70 DAS	80 DAS	90 DAS	Mean	70 DAS	80 DAS	90 DAS	Mean	70 DAS	80 DAS	90 DAS	Mean	Mean	check (%)		
<i>Trichoderma viride</i>	1.00	24.3(29.5)	42.7(40.8)	76.5(61.0)	47.8(43.8)	25.1(30.0)	42.9(40.9)	77.3(61.6)	48.4(44.1)	48.1	0.8					
<i>Pseudomonas fluorescens</i>	1.00	23.7(29.2)	42.1(40.5)	76.3(60.9)	47.4(43.5)	24.8(29.9)	42.7(40.8)	77.1(61.4)	48.2(44.0)	47.8	1.6					
Salicylic Acid	0.25	13.9(21.9)	33.6(35.4)	57.3(49.2)	34.9(36.2)	11.2(19.6)	28.8(32.5)	54.7(47.7)	31.6(34.2)	33.2	31.5					
Borax	0.50	16.3(23.8)	36.0(36.9)	69.1(56.2)	40.4(39.5)	16.8(24.2)	33.9(35.6)	63.7(53.0)	38.1(38.1)	39.3	19.1					
K <sub>2</sub> SO <sub>4</sub>	1.00	18.1(25.2)	37.6(37.8)	71.2(57.5)	42.3(40.6)	18.9(25.8)	35.2(36.4)	67.7(55.4)	40.6(39.6)	41.5	14.6					
CaSO <sub>4</sub>	1.00	15.2(23.0)	35.2(36.4)	65.6(54.1)	38.7(38.5)	15.2(23.0)	31.2(34.0)	58.9(50.2)	35.1(36.3)	36.9	24.0					
Metalaxyl0.1%	0.10	12.8(21.0)	29.6(33.0)	52.0(46.2)	31.5(34.1)	9.6(18.1)	23.5(29.0)	46.9(43.2)	26.7(31.1)	29.1	40.1					
Metalaxyl0.2%	0.20	12.5(20.7)	29.3(32.8)	51.7(46.0)	31.2(34.0)	9.3(17.8)	23.2(28.8)	46.7(43.1)	26.4(30.9)	28.8	40.7					
Metalaxyl0.3%	0.30	12.3(20.5)	29.1(32.6)	51.5(45.8)	30.9(33.8)	9.1(17.5)	22.9(28.6)	46.4(42.9)	26.1(30.7)	28.5	41.2					
KCl	1.00	20.0(26.6)	39.2(38.8)	72.0(58.1)	43.7(41.4)	20.0(26.6)	37.3(37.7)	71.5(57.7)	42.9(40.9)	43.3	10.7					
ZnSO <sub>4</sub>	0.50	21.3(27.5)	41.1(39.9)	73.9(59.3)	45.4(42.4)	22.4(28.3)	39.2(38.8)	75.5(60.3)	45.7(42.5)	45.6	6.1					
Check	-	24.8(29.9)	43.2(41.1)	76.8(61.2)	48.3(44.0)	25.3(30.2)	43.5(41.3)	77.6(61.8)	48.8(44.3)	48.5	0.0					
SE <sub>±</sub>	-	0.48	0.86	0.65	-	0.73	0.86	1.70	-	-	-					
CD 5%	-	1.39	2.47	1.87	-	2.10	2.48	4.89	-	-	-					

Values in parenthesis are angular transformed values

DAS= Days after sowing

17. In non-conventional chemicals, salicylic acid was found superior followed by calcium sulphate then others which reduce average number of pustules 37.95 % and 32.07 % in comparison with the control, respectively and minimum reduction in number of pustules were recorded in zinc sulphate at 0.50 % (7.4 %) followed by Borax 0.50 % (15.8%).

The per cent disease index in cv. RH-749 was recorded at 70, 80 and 90 days after sowing of the mustard. All the treatments were found significantly better over the check (Table 3). Among all the treatments Metalaxyl 0.3% found superior which reduce 41.21 per cent disease in comparison with the control. However, the different concentrations of Metalaxyl were found at par with each other at 70, 80 and 90 days after sowing during 2015-16 and 2016-17. In bio agents, Both *T. viride* and *P. fluorescens* were found non-significant at 70, 80 and 90 days after sowing during 2015-16 and 2016-17 in comparison with the control. Among abiotic agents, salicylic acid at 0.25% reduces disease significantly in 2015-16 and 2016-17. Salicylic acid 0.25% reduces 31.5% white rust disease over the control followed by calcium sulphate at 1.0% which reduces disease 24.0%. Zinc sulphate at 0.50% was found least effective abiotic chemical which reduce 6.1% disease.

Several plant defense activators in the management of *A. candida* has been used in mustard (Tirmali and Kolte, 2012). They found calcium sulphate, potassium chloride, potassium sulphate, zinc sulphate and borax significantly superior effective in reduction the pustules size of white rust on the mustard leaves with comparison to control. Sharma and Kolte (1994) reported that potassium fertilized plants exhibited 30 to 45 per cent less disease severity of Alternaria blight based on the number and size of the spots, average disease index on leaf and pods. Tewari (1991) found that foliar application of the calcium reduce the per cent disease severity of Alternaria blight in rapeseed. Antonova *et al.* (1984) and Dixon *et al.* (1987) reported that boron application in the cabbage increase resistance to club root.

## References

Antonova TS, Zaichlor VE and Kalinchenlw TV. 1984. Role of calcium in the resistance of the sunflower anthodium to gray root. *Sci Skokhozyaistvernaya Biologiya* **11**: 67-68.

Becot S, Pajot E, Corre D, Monot C and Silne D. 2000. Phytogard ( $K_2PHO_3$ ) induces localized resistance in cauliflower to downy mildew of crucifers. *Crop Protect* **19**: 417-425.

Benhamon N and Garand C. 2001. Cytological analysis of defense related mechanisms in pea root tissues in response to colonization by nonpathogenic *Fusarium oxysporum* Fo47. *Phytopathol* **91**: 730-740.

Biswas C, Singh R and Kumar PV. 2011. Dynamics of white rust disease in mustard (*B. juncea*) in relation to date of sowing and weather parameters. *Indian J Agril Sci* **81**: 1187-1190.

Cohen Y. 1994. 3-Aminobutric acid induces systemic resistance against *Peronospora tabacina*, *Physiol Mol Plant Pathol* **44**: 273-288.

De Oliveira DE, Seurinck J, Inze D, Van Montague M and Botterman J. 1990. Differential expression of five *Arobidopsis* genes encoding glycine-rice protein. *The Plant Cell* **2**: 427-436.

Dixon GR, Naiki T, Webster A and Wilson F. 1987. Integrated use of boron calcium cyanamide and nitrogen for control of club root (*Plasmodiophora brassicae*), *In: Crop Protection in Northern Britan*: 399-404.

Droly S, Vinokur V, Wiess B, Cohen L, Daws A, Goldschmidt EE and Porat R. 2002. Induction of resistance to *Pennicilium digitatum* in grape fruit by the yeast bio-control agent *Candida oleophila*. *Phytopathol* **92**: 93-399.

Durrant WE and Dong X. 2004. Systemic acquired resistance. *Ann Rev Phytopathol* **42**: 185-209.

Gorlach J, Volrath S, Oostendorp M, Kogel KH, Beckhove U, Staub T, Ward E, Kessmann H and Ryals J. 1996. Benzothiadiazole, a novel class of inducers of systemic acquired resistance, activates systemic resistance in wheat. *Plant Cell* **8**: 629-643.

Graham JH and Leite RP. 2004. Lack of control of citrus canker by induced systemic resistance compounds. *Plants Dis* **88**: 745-750.

Hammerschmidt R. 1999. Induced disease resistance: how do in-duced plants stop pathogens. *Physiol Mol Plant Pathol* **55**: 77-84.

Holopainen JK, Heijari J, Nerg AM, Vuorinen M and Kainulainen P. 2009. Potencial for the use of exogenous chemical elicitors in disease and insect pestmanagement of conifer seedling production. *Open For Sci J* **2**: 17-24.

Howell CR, Hanson LE, Spitanovic RD and Puckhaber CS. 2000. Induction of terpenoid synthesis in cotton roots and control of *Rhizoctonia solani* by seed treatments with *T. virens*. *Phytopathol* **90**: 248-252.

Kaur A and Kolte SJ. 2001. Protection of mustard plants against staghead phase of white rust by Foliave treatment with benzothiadiazole-An activator of plant defense system. *J Mycol Pl Pathol* **31**: 133-138.

- Kobayashi A, Tai A, Hiroshi K and Kazuyoshi K. 1993. Elicitor-active oligosaccharides from algal laminarin stimulate the production of antifungal compounds in alfalfa. *Z Naturforsch* **48**: 575-579.
- Liu L, Kloepper JW and Tuzun S. 1995. Induced systemic resistance in cucumber against bacterial angular spot by plant-growth promoting rhizobacteria. *Phytopathol* **85**: 843-847.
- Macmillan RT, Graves W and Mc Laughlin HJ. 2000. Key Plex effect on Sigatoka of banana. *PWC Florida State Hort Soc* **112**: 192-193.
- Matheron ME and Porchas M. 2002. Suppression of Phytophthora root and crown rot on pepper plants treated with Acibenzolar-SMethyl. *Plant Dis* **86**:292-297.
- Mathur P, Sharma E, Singh SD, Bhatnagar AK, Singh VP and Kapoor R. 2013. Effect of elevated CO<sub>2</sub> on infection of three foliar diseases in oilseed *B. juncea*. *J Plant Pathol* **95**: 135-144.
- Narusaka X, Naraska M, Horio T and Ishii H. 1999. Comparison of local and systemic induced resistance in cucumber plants treated with benzothiadiazoles or salicylic acid. *Plant Cell Physiol* **40**:388-395.
- Omidi H. 2010. Changes of proline content and activity of antioxidative enzymes in two canola genotype under drought stress. *Amr J Plant Physiol* **5**:338-349.
- Ozeretskovskaya OL and Vasyukova NI. 2002. The use of elicitors for protection of cultured plants demands caution. *Appl Biochem Microbiol* **38**:277-279.
- Pajot E, Corre D Le and Silue D. 2001. Phytogard and DL- $\beta$ -amionobutyric acid (BABA) induced resistance to downy mildews (*Bremia lactucae*) in lettuce (*Lettuce sativa*). *Eur J Plant Pathol* **107**: 861-864.
- Panase VG and Sukhatme PV. 1985. Statistical Methods for Agricultural Workers, 4th ed., ICAR, New Delhi, 347 p.p
- Romero AM, Kousik CC and Ritchie DF. 2001. Resistance to bacterial spot in bell pepper induced by acibenzolar-5-methyl. *Plant Dis* **85**: 189-194.
- Ryals J, Uknes S and Ward E. 1994. Systemic acquired resistance. *Plant Physiol* **104**: 1109-1112.
- Safari S, Soleimani MJ, Mohajer A and Fazlikhani L. 2013. Possible structure-activity profile of salicylate derivatives: their relationship on induction of systemic acquired resistance. *J Agril Technol* **9**: 1215-1225.
- Sharma SR and Kolte SJ. 1994. Effect of soil applied NPK fertilizers on severity of black spot disease (*Alternaria brassicae*) and yield of oil seed rape. *Plant Soil* **167**:313-320.
- Singh US, Dought KJ, Nashaat NI, Bennet RN and Kolte SJ. 1999. Induction of systemic resistance to *Albugo candida* in *B. juncea* by pre or co-inoculation with an incompatible isolate. *Phytopathol* **89**: 1226-1232.
- Spletzer ME and Enyedi AJ. 1999. Salicylic acid induces resistance to *Alternaria solani* in hydroponically grown tomato. *Phyto-pathol* **89**: 722-727.
- Surekha CH, Neelapu NNR, Siva Prasad B and Sankar GP. 2014. Induction of defense enzymes and phenolic content by *T. viride* in *Vigna mungo* infested with *Fusarium oxysporum* and *Alternaria alternate*. *Intl J Agril Sci Res* **4**: 31-40.
- Tewari JP. 1991. Current understanding of resistance to *A. brassicae* in crucifers. Proc. *GCRIC*, 8<sup>th</sup> Intl. Rapeseed Cong. July 9-11, Saskatoon, Canada **2**: 471-476.
- Tirmali AM and Kolte SJ. 2012. Induction of host resistance in mustard with non-conventional chemicals against white rust (*A. candida*). *J Pl Dis Sci* **7**: 27-31.
- Verhagen BMW, Glazebrook JT, Chang HS and Van Loon LC. 2004. The transcriptome of Rhizobacteria induced Systemic resistance in *Arabidopsis*. *Mol Plant Microbe Interact* **17**: 985-908.
- Vishwanath C, Kolte SJ, Singh MP and Awasthi RP. 1999. Introduction of resistance in mustard (*B. juncea*) against Alternaria black spot with an avirulent *Alternaria brassicae* isolate-D. *Euro J Plant Pathol* **105**: 217-220.
- Wei C, Laby RJ, Zumoff CH, Bauer HW, He SY, Colmer A and Beer SV. 1992. Harpin, elicitor of the hypersensitive response produced by the plant pathogen *Erwinia amylovora*. *Sci* **257**:1-132.
- Wheeler BEJ. 1969. An Introduction to Plant Diseases. John Wiley and Sons, London, UK.
- Yalpani N and Raskin I. 1993. Salicylic acid: a systemic signal in induced plant resistance. *Trends Microbiol* **1**:88-92.
- Zhao J, Davis LC and Verporte R. 2005. Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnol Adv* **23**: 283-333.