



## Genetic diversity among different isolates of *Sclerotinia sclerotiorum* in North Iran

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### Abstract

*Sclerotinia sclerotiorum* (Lib.) de Bary is a cosmopolitan, homothallic fungus, and is the most important causal agent of stem rot diseases in field crops of Iran. During 2011-2012, a total of 52 isolates of the fungus were collected from infected rapeseed plants (*Brassica napus* cultivar Hiola 401) from several fields of Northern Iran. The genetic diversity of *S.sclerotiorum* populations were assessed using Simple Sequence Repeat fingerprinting (SSR). By using five SSR primers, 27 haploid groups with 14 alleles polymorphism were detected. A high level of genetic diversity 52% was observed between isolates. The results showed that there were possibly high rates of out crossing, as well as, evolutionary potential within 52 isolates obtained from different geographical locations. The variability found within closely related isolates demonstrates the effectiveness of SSR marker in identifying genetic diversity. This is the first report on genetical diversity in *S. sclerotiorum* populations in Mazandaran province in Northern Iran.

**Key words:** Genetic diversity, microsatellites, *Sclerotinia sclerotiorum*

### Introduction

*Sclerotinia sclerotiorum* (Lib.) de Bray an ascomycetous fungus, causing white mould disease on more than 400 agricultural and wild plant species has a worldwide geographical distribution (Purdy, 1979; Boland and Hall, 1994). *Sclerotinia sclerotiorum* is a homothallic fungus which disperses by either airborne ascospores, or by soil-borne sclerotia. The sclerotia are the resting propagules that germinate to produce either hyphae or apothecia (Abawi and Grogan, 1975; Adams and Ayers, 1979; Willets and Wong, 1980; Hao *et al.*, 2003). *Sclerotinia* rot is the most destructive disease of canola, especially in favourable climatic conditions, in the northern flats of the Caspian Sea (Pakdaman and Mohammadi, 2007). In Iran, the evaluation of several cultivars and genotypes of rapeseed against *Sclerotinia* rot has shown different levels of tolerance (Zhao *et al.*, 2004).

Control strategies must target a population, instead

of an individual, if they are to be effective (McDonald, 1997). In plant-pathogen interactions, The development of new pathogenic races, and the breakdown of host resistance are the limiting factors in resistance deployment against plant diseases. The pathogen's life history characteristics and evolutionary potential are major factors leading to the pathogen overcoming host resistance (Gaggiotti *et al.*, 1999; Coletta-Filho and Machado, 2002; McDonald and Linde, 2002). Therefore, major efforts should be focused not only in understanding the genetic structure of the fungal populations, But also to determine how populations will evolve in response to different control strategies (McDonald, 1997).

Two independent mechanisms, mycelial compatibility group (MCG) and DNA fingerprinting, are known to differentiate *S. sclerotiorum* populations. DNA fingerprinting technique can also be used to distinguish closely related fungal isolates. Southern hybridisation of restriction digested whole genomic

DNA to a cloned probe containing a 4.5 kb repeated dispersed element of nuclear DNA from *S. sclerotiorum* has been used previously (Kohn *et al.*, 1991, Kohli *et al.*, 1995; Cubeta *et al.*, 1997). Microsatellites (Litt and Luty, 1989), also known as simple sequence repeats (SSRs), or short tandem repeats (STRs), are the smallest class of simple repetitive DNA sequences (Tautz *et al.*, 1986; Mc-Donald and Potts, 1997), that are widely dispersed and evenly distributed in the genome of eukaryotes, has also been used to study intraspecific genetic variability within populations (Gaggiotti *et al.*, 1999; Singh and Kohn, 2001). The aim of this present research was to investigate the genetic structure within the oilseed rape population of *S. sclerotiorum* in the Mazandaran province of Iran based on molecular markers.

## Materials and Methods

### Pathogen isolates

*S. sclerotiorum* isolates were collected from 11 rapeseed fields in Mazandaran province of northern Iran during 2011-2012 crop season. Sclerotia from infected plants were removed and a single sclerotium was selected as an isolate. Sclerotia were surface sterilized for 1 min. in 70% ethanol, or 2 min. in 2.5% sodium hypochlorite, rinsed in sterilized-distilled water, plated on potato dextrose agar medium (PDA), and incubated at 22 °C for two days. Each isolate was purified by transferring the single hyphal tip onto the fresh medium, and the generated sclerotia were stored at -20°C until use (Sharma *et al.*, 2013; Atallah *et al.*, 2004).

### DNA Extraction

Isolates were grown on PDA and incubated at 21- 24°C in darkness for 5 days. Mycelia were harvested by vacuum filtration, lyophilised, and stored at -20°C. Fifteen to 20 mg of dried mycelia were powdered, and transferred to 1.5 Eppendorf tubes for DNA extraction following the method of Liu *et al.* (2000), with the slight modifications. The DNA pellets were dissolved in 30 µl of 1X TE (10 mM Tris-HCl, 1 mM EDTA), and stored at 5°C (Liu *et al.*, 2000).

Three sets of microsatellite primers including CATA<sub>25</sub>, CA<sub>9</sub>, and TACA<sub>10</sub> were used in this study

(Sirjusingh and Kohn, 2001). Gels were stained with ethidium bromide, visualized under UV light, and digitally documented with the gel documentation UVP-V system. The gel was run at 90 W for 90 min (Nicholson *et al.*, 1997). All polymorphic alleles were identified from each microsatellite primer combination, and bands representing alleles were scored as present (1) or absent (0). Nei's genetic distance matrix (Nei and Li, 1979) was prepared, and bootstrap analysis with 2000 replications, was performed to generate a dendrogram of unweighted pair-group mean analysis (UPGMA; Sokal and Michener, 1958) using the TREECON 1.3b program (Van de Peer and de Wachter, 1994).

### Polymorphism Information Content (PIC)

The polymorphism information content (PIC) determining the frequency of alleles polymorphism in the gene locus of the population, was calculated as follows :

$$PIC = 1 - \sum_{i=1}^n p_i^2$$

Where  $p_i$  was the relative frequency of  $i$ th alleles

## Results and Discussion

### Genetic diversity among the *Sclerotinia sclerotiorum* isolates

In 52 isolates the microsatellite primers exhibited 14 clear polymorphic alleles. The number of polymorphic alleles per locus ranged from 3 to 6 (table 1). The genetic relationships among the isolates was determined by using a separate matrix to the data from the 27 polymorphic alleles, and the 52 isolates were then clustered (Figure 1).

The polymorphism information content (PIC) for each locus ranged from 0.388 for (CA)<sub>9</sub> to 0.781 for (CATA)<sub>25</sub> (table 1) . Therefore, the Locus (CATA)<sub>25</sub> contained the most identified alleles and the highest PIC Although a high level of diversity was observed between the clusters, some isolates within clusters were identical for all the microsatellite markers. For instance, isolates 41 and 42 had similar alleles for all the microsatellite markers. Overall, 27 distinct isolates were identified among the 52 isolates.

In the present study, it was discovered that

Table 1: Microsatellite and polymorphic loci, and polymorphism information content (PCR) for *S. sclerotiorum*

Locus (Accessions No.)	Repeat motifs	Primer sequence (5-	PIC	Allele Number	Previous reports	Other species
12-2 AF377906	(CA) <sub>9</sub>	CGATAATTTCCCCTCACTTGC GGAAGTCCTGATATCGTTGAGG	0.388	3	4 <sup>a</sup> 2 <sup>b</sup>	No
106-4 AF377921	(CATA) <sub>25</sub>	TGCATCTCGATGCTTGAATC CCTGCAGGGAGAAACATCAC	0.781	6	10 <sup>a</sup> 3 <sup>b</sup>	Yes
55-4 AF377918	(TACA) <sub>10</sub>	GTTTTTCGGTTGTGTGCTGG GCTCGTTCAAGCTCAGCAAG	0.611	5	7 <sup>a</sup> 4 <sup>-b</sup>	Yes

Notes: <sup>a</sup>Sirjusingh and Kohn, 2001      <sup>b</sup>Atallah *et al.*, 2004

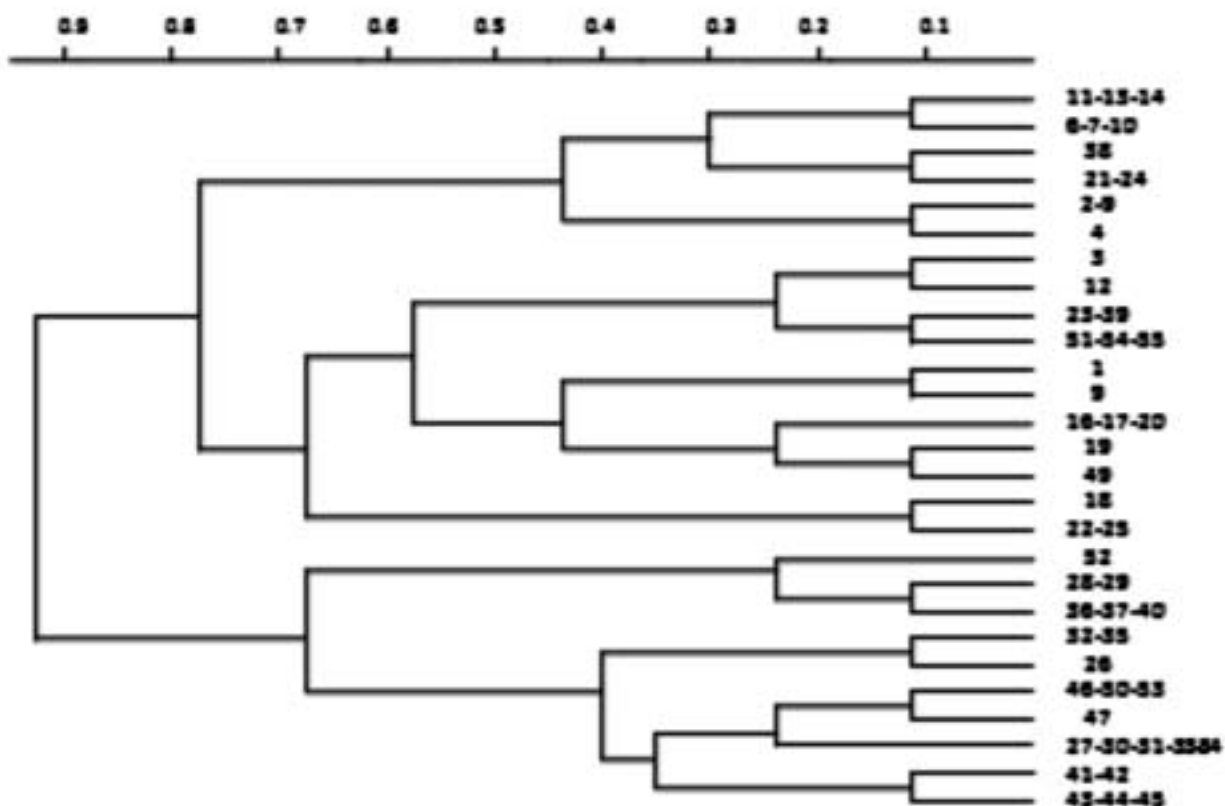


Fig.1. Unweighted pair-group mean analysis dendrogram of genetic distance among the 65 fungal isolates based on Jaccard's coefficients.

microsatellite markers were very efficient in identifying genetic variation among the isolates. Three of these marker sets identified polymorphism among the Iranian isolates. When our findings were compared with the previous work (Atallah *et al.*, 2004). Four new alleles, one at (TACA)<sub>10</sub> and (CA)<sub>9</sub>, and three at (CATA)<sub>25</sub>, were discovered (Atallah *et al.*, 2004; Singh and Kohn, 2001).

Results of our present study showing presence of 27(S) different clones (haplotypes) among the 52(N) isolates strongly suggests that a high degree of genetic variability (S/N%=52%) exists in the *S. sclerotiorum* population in the mazandaran province of northern Iran. From Australia Sexton and Howlett (2004) also reported presence of genotypic diversity, ranging from 36-80%, amongs

*S. sclerotiorum* isolates collected from four oilseedrape fields. From Newzealand Carpenter *et al.* (1999) have also reported shared microsatellite haplotypes amongst *S. sclerotiorum* isolates collected from different fields.

Shared haplotypes among populations indicate either considerable gene flow (exchange of haplotypes among populations) and the populations represent the same founder population, shared haplotypes among populations could also be the result of sexual reproduction, i.e. inbreeding or outcrossing among similar genotypes. The resultant ascospores would be genetically similar and appear clonal. Because ascospores can be dispersed over long distances (Cubeta *et al.*, 1997), it is not surprising to find shared clonal genotypes among several regions. Shared microsatellite haplotypes among populations have also been reported from Australia and North America (Sexton and Howlett, 2004; Atallah *et al.*, 2004). The isolates in the present study, were collected from several fields in northern Iran where diverse crops are cultivated, and where environmental conditions favour sexual recombination within *S. sclerotiorum*. Oilseed rape is a new crop to this area and the high level of polymorphism may reflect the movement of *S. sclerotiorum* ascospores onto rapeseed crop from several wild host plants.

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### References

- Abawi, GS and Grogan, RG. 1975. Source of primary inoculum and effects of temperature and moisture on infection of beans by *Whetzelinia sclerotiorum*. *Phytopath* **65**: 300-309.
- Atallah, Z, Larget, B and Johnson, D. 2004. High genetic diversity, phenotypic uniformity and evidence of outcrossing in *Sclerotinia sclerotiorum* in the Columbia basin of Washington state. *Phytopath* **94**: 737-742.
- Adams, PB and Ayers, WA. 1979. Ecology of *Sclerotinia* species. *Phytopath* **69**: 896-898.
- Boland, G and Hall, R. 1994. Index of plant hosts of *Sclerotinia sclerotiorum*. *Can J Plant Pathol* **2**: 93-108.
- Carpenter, MA, Frampton, C and Stewart, A. 1999. Genetic variation in New Zealand populations of the plant pathogen *Sclerotinia sclerotiorum*. *N Z J Crop Hort Sci.* **27**: 13-21.
- Cubeta, MA, Cody, BR, Kohli, Y and Kohn, L. 1997. Clonality in *Sclerotinia Sclerotiorum* on infected cabbage in eastern North Carolina. *Phytopath.* **87**: 1000-1004.
- Coletta-Filho, HD and Machado, MA. 2002. Evaluation of the genetic structure of *Xylella fastidiosa* populations from different *Citrus sinensis* varieties. *Appl Environ Microbiol* **68**: 3731-3736.
- Gaggiotti, OE, Lange, O, Rassmann, K and Gliddon, C. 1999. A comparison of two indirect methods for estimating average levels of gene flow using microsatellite data. *Mol Ecol* **8**: 1513- 1520.
- Hao, JJ, Subbarao, KV and Duniway, JM. 2003. Germination of *Sclerotinia minor* and *S. sclerotiorum* sclerotia under various soil moisture and temperature combinations. *Phytopath* **93**: 443-450.
- Kohli, Y, Brunner, LJ, Yoel, H, Milgroom, MG, Anderson, JB, Morrall, RAA and Kohn, LM. 1995. Clonal dispersal and spatial mixing in populations of the plant pathogenic fungus, *Sclerotinia sclerotiorum*. *Mol Ecol* **4**: 69-77.
- Kohn, LM Stasoviski, E Carbone, I Royers, J and Anderson, J 1991. Mycelial incompatibility and molecular markers to identify genetic variability in field populations of *Sclerotinia sclerotiorum*. *Phytopath* **81**: 480-485.
- Litt, M and Luty, JM 1989. A hypervariable microsatellite revealed by in vitro amplification of a dinucleotide repeat within the cardiac muscle actin gene. *Amer J Hum Genet* **44**: 397-401.
- Liu, D. Coloe, S. Baird, R. and Pedersen, J. 2000. Rapid mini-preparation of fungal DNA for PCR. *J Clinical Microbio* 471pp.
- McDonald, BA 1997. The population genetics of fungi: Tools and Techniques. *Phytopath* **87**: 448-453.

- McDonald, BA and Linde, C. 2002. The population genetics of plant pathogens and breeding strategies for durable resistance. *Euphytica* **124**: 163-180.
- McDonald, DB and Potts, WK 1997. *DNA microsatellites as genetic markers for several scales*. In: Mindell D.P. (ed.), Avian molecular evolution and systematics. *Academic Press, San Diego* 29-49.
- Nei, M and Li, WH. 1979. Mathematical models for studying genetic variation in terms of restriction endonucleases. *Proc Natl Acad Sci, USA* **76**: 5268-5273.
- Nicholson, P, Rezanoor, HN Simpson, DR and Joyce, D. 1997. Differentiation and quantification of the cereal eyespot fungi *Tapesia yallundae* and *Tapesia acuformis* using a PCR assay. *Plant Pathol* **46**: 842-856.
- Pakdaman, BS and Mohammadi Goltapeh, E. 2007. In vitro studies on the integrated control of rapeseed white stem rot disease through the application of herbicides and *Trichoderma* species. *Pak J Biotech Sci* **10**: 7-12
- Purdy, L. 1979. *Sclerotinia sclerotiorum* – history, diseases and symptomatology, host range, geographic distribution, and impact. *Phytopath* **69**: 875-880.
- Sexton, AC and Howlett, BJ. 2004. Microsatellite markers reveal genetic differentiation among populations of *Sclerotinia sclerotiorum* from Australian canola fields. *Curr Genet* **46**: 357-365.
- Sharma, P, Meena, PD, Kumar, S and Chauhan JS. 2013. Genetic diversity and morphological variability of *Sclerotinia sclerotiorum* isolates of oilseed Brassica in India. *African J Microbiol Res* **7**: 1827-1833.
- Singh, SC and Kohn, LM 2001. Characterization of microsatellites in the fungal plant pathogen, *Sclerotinia sclerotiorum*. *Mol Ecol Notes* **1**: 267-269.
- Sokal, RR and Michener, CD. 1958. A statistical method for evaluating systematic relationships. *Univ Kansas Sci Bull* **38**: 1409-1438.
- Van de Peer, Y and de Wachter, R. 1994. TREECON for Windows: a software package for the construction and drawing of evolutionary trees for the Microsoft Windows environment. *Comput Appl Biosci* **10**: 569-570.
- Willems, H. and Wong, J. 1980. The biology of *Sclerotinia sclerotiorum*, *S. trifoliorum* and *S. minor* with emphasis on specific nomenclature. *Bot Rev* **46**: 101-165.
- Tautz, D, Trick, M and Dover, GA. 1986. Cryptic simplicity in DNA is a major source of genetic variation. *Nature* **322**: 652-656.
- Zhao, J, Peltier, AJ, Meng, J, Osborn, TC and Grau, CR. 2004. Evaluation of *Sclerotinia* stem rot resistance in oilseed *Brassica napus* using a petiole inoculation technique under greenhouse conditions. *Plant Dis* **88**: 1033-1039.